have significant implications in areas where charge transport in nanostructured materials and devices is becoming increasingly important, such as ion conductors, photovoltaics or electroluminescence, for which the present extended amphiphilic dendrons may provide an advanced molecular design concept.

References and Notes

41. This work was supported by the Postdoctoral Fellowship Program of Korea Science and Engineering Foundation (KOSEF), the NSF (DMR-0312913), and the Cornell Center for Materials Research (CCMR), a Materials Research and Science and Engineering Center (MRSEC) of the NSF (DMR-0079992). Financial support of Philip Morris, USA, is gratefully acknowledged. The SAXS x-ray facility is supported by Department of Energy Biological and Environmental Research (BER) grant DE-FG02-97ER62243. This work made use of the CCMR Hudson mesoscale facility, supported through the NSF MRSEC Program (DMR-0079992). We thank H. J. Räder and S. Türk (Max-Planck Institute for Polymer Research) for the MALDI-TOF measurements on the extended dendrimers and S. Mahajan (Cornell University) for help with PEO synthesis.
agents,” analogous to tRNAs) (Figs. 2B and 3). Each reagent oligonucleotide complemented one of three unique coding regions in the template sequence. The reagent oligonucleotides were biotinylated to allow products from each DNA-templated step (5a, 6a, and 7a) to be purified by capture with, and release from, streptavidin-linked magnetic beads (8) (Fig. 2, A and B). This direct selection for bond formation facilitates multistep synthesis by enabling the one-pot purification of products independent of their structure.

The diol group in the captured product of the third DNA-templated reaction (7a) was oxidatively cleaved with NaIO$_4$ to reveal an aldehyde. The phosphonium group was then deprotonated by elevating the pH of the buffer to 8.5, which induced Wittig olefination and macrocyclization. Because macrocyclization results in the cleavage of the reagent oligonucleotide-product bond, the desired macrocyclic fumaramide (8a) self-eluted in pure form from the streptavidin-linked magnetic beads (Fig. 2, A and B). The generality of this reaction was examined by assaying the macrocyclization of 11 molecules related to 7a (fig. S1) (13). Macrocyclization was efficient for a wide variety of precyclized structures (60 to 90% yields with no contaminating uncyclized material). Control reactions lacking NaIO$_4$ confirmed that the cyclization reaction required the presence of an aldehyde group (fig. S1) (13). The progress of each DNA-templated step during the transformation of 1a to 8a was followed by denaturing polyacrylamide gel electrophoresis (PAGE) (Fig. 2C) and by matrix-assisted laser desorption/ionization–time-of-flight (MALDI-TOF) mass spectrometry (Fig. 4A) after endonuclease-catalyzed removal of all but seven nucleotides at the 5’ end of the template. The presence of

Fig. 1. Scheme for the translation, selection, and amplification of libraries of DNA templates encoding synthetic small molecules. When the number of different possible library structures approaches or exceeds the number generated, template diversification after selection can be added to evolve the pool of synthetic molecules toward structures possessing desired properties. X is a starting material common to all library members.

Fig. 2. (A) DNA-templated macrocycle library synthesis scheme. R is NHCH$_3$ where R is NHCH$_3$ or tryptamine; Ar, –(p-C$_6$H$_4$)–. The macrocyclization reaction is confirmed to give predominantly trans alkene stereochemistry for one library member (fig. S4) (13) but may give other outcomes for different macrocyclic structures. (B) Template and reagents used in the DNA-templated synthesis of 8a. (C) Denaturing PAGE of each step in the DNA-templated synthesis of 8a. Lane h is the product of a DNA-templated thiol addition to the product shown in lane g, confirming the formation of the fumaramide group during macrocyclization.
the electrophilic fumaramide group in the final product was confirmed by mass spectrometry and by its ability to accept a thiol nucleophile during a DNA-templated conjugate addition (Figs. 2C and 4A). Macrocycle 8a (expected mass is 2908.8 daltons; observed mass is 2910.4 ± 6 daltons) was synthesized with high purity in 1 to 5% overall yield for three DNA-templated steps, the macrocyclization, and all associated purifications. To verify that the reaction conditions do not induce epimerization of the amino acid–derived chiral centers, we performed large-scale amine acylation reactions of analogous non–DNA-linked substrates under conditions that mimicked steps 1 to 3. The stereochemical integrity of the resulting products was confirmed by comparison to authentic diastereomeric standards (Figs. S2 and S3) (13). In addition, exposure of a non–DNA-linked version of 7 to the above macrocyclization conditions provided the corresponding macrocycle (a non–DNA-linked version of 8) containing a predominantly trans alkene (14) by nuclear magnetic resonance analysis (Figs. S4 and S5) (13).

In addition to the multistep DTS of one synthetic small molecule, implementation of the scheme in Fig. 1 requires that DTS proceed in a sequence-specific manner in a library format in which multiple templates and multiple reagents are present in the same solution. Although DNA-templated reactions have been shown to be sequence-specific (5, 9), library-format DTS to generate multistep small-molecule products has not been previously achieved.

We chose unique template “codons” to encode four or five reactants for each of the three DNA-templated steps in the macrocycle synthesis (13 codons total) (Fig. 3A). Each of the 13 codons was assigned to encode a different building block. The building blocks were chosen to include diverse functionalities, stereochemistries, and backbone lengths (Fig. 3A). Four different templates (1a to 1d) (Fig. 3B), each containing three codons, were prepared such that the maximum number of 12 different codons (complementing reagents 2a to 2d, 3a to 3d, and 4a to 4d) were represented within the four templates. The corresponding reagents, each consisting of an amino acid building block conjugated through the linkers shown in Fig. 2A to a decoding DNA “anticodon,” were also prepared (Fig. 3A).

We tested the sequence specificity of DTS in the presence of multiple reagents by exposing template 1a, 1b, 1c, or 1d separately to a mixture of all step 1 reagents except the reagent complementing the step 1 codon present in each template. As a positive control, each of the four templates was also separately reacted with its complementary step 1 reagent. These two DNA-templated reactions were repeated for each of the four step 2 codons and for each of the four step 3 codons. In contrast with the positive control, the reaction lacking the complementary reagent in all 12 cases did not generate significant product (Fig. 3C). These results indicate that templates do not react with mismatched reagents under the conditions in Fig. 2, even in the absence of complementary reagents.

To examine the sequence specificity of true library-format DNA-templated synthesis involving multiple templates and multiple reagents in a single solution, we reacted templates 1a to 1d in one solution with the four step 1 reagents (2a to 2d). After reagent-linker cleavage (Fig. 2, A and C), the solution containing the step 1 products was reacted with the four step 2 reagents (3a to 3d), and the resulting purified products were then reacted with the four step 3 reagents (4a to 4d) before undergoing Wittig macrocyclization.

In all cases, the major products observed by MALDI-TOF mass spectrometry after each step consisted of all four of the sequence-programmed products (Fig. 4B). If the 12 reagents used to synthesize 8a to 8d reacted with templates randomly, rather than in a sequence-programmed manner, up to 64 different macrocycles would have been synthesized rather than the exclusive formation of the four observed products. The faithful translation of four DNA templates (1a to 1d) into four sequencel-programmed macrocyclic fumaramides (8a to 8d) indicates a one-to-one correspondence between the DNA sequence that enters the above process and the structure of the resulting macrocycle.

After developing a robust multistep DNA-templated macrocycle synthesis and establishing sequence specificity of each template-reagent combination used in the macrocycle library synthesis. The variable regions within each anticodon are underlined. The NlaI cleavage site within template 1e is shown in lower case. (B) Representative DNA templates used in macrocycle library synthesis. Templates 1a to 1e (R is NHCH₃ or tryptamine) collectively call for each of the reagents in (A). (C) Denaturing PAGE analysis confirming the results of the library synthesis. The template and reagent(s) were combined under the conditions shown in Fig. 2A, and the reactions were analyzed before reagent-linker cleavage. Products appear as bands of higher molecular weight above templates.
the sequence specificity of library-format DNA-templated reactions, we prepared a library of 65 templates (1) that contained all 64 possible combinations of the four codons at each coding region and a 65th template (1e) that uniquely contained a step 1 codon encoding a phenyl sulfonamide building block (Fig. 3A). Because carbonic anhydrase is known to bind phenyl sulfonamides with high affinity [dissociation constant $K_d \approx 1 \text{nM}$ (15)], the macrocycle encoded by the 65th template serves as a positive control to evaluate the ability of a DNA-templated small molecule library to be selected in vitro for target protein affinity (see below).

The single-solution library of 65 equimolar DNA templates was translated into 65 corresponding macrocyclic fumaramides through the scheme shown in Fig. 2A. Each of the three coupling steps was executed in a single solution containing all 65 templates (typically 500 pmol total) and all five (step 1) or four (steps 2 and 3) reagents as described above. Denaturing PAGE analysis of each library synthesis step indicated yields similar to those of the single-template and four-template cases. MALDI-TOF mass spectrometry was used to observe the formation of the four major step 1 small-molecule products (all but the product encoded by 1e, which is expected to be 16-fold less abundant than the four major step 1 products). After step 2, mass spectrum peaks consistent with the presence of all mass-resolvable step 2 small-molecule products were also observed (Fig. 4C).

After step 3 and macrocyclization, exposure of the completed 65-member library to a DNA-linked thiol reagent efficiently (84% yield) converted the library to higher molecular weight species. This result is consistent with the formation of the fumaramide group during the macrocyclization (Fig. 4D). Beginning with 0.1 to 3 nmol of starting template (1), the multistep DNA-templated library synthesis described above provided sufficient final product to undergo many in vitro selections (10) for library members with protein binding properties. Taken together, these results represent the translation of a library of DNA templates into a library of corresponding synthetic molecules.

Each member of a DNA-templated synthetic library of the type described in Fig. 1 is associated with an amplifiable DNA strand that not only encodes but has actually directed that molecule’s synthesis. DNA-templated libraries, like libraries made by DNA display (16), are conceptually analogous to genetically encoded protein libraries (2, 4) except that the structures generated are not limited to those that can be biosynthesized by the ribo-

---

**Fig. 4.** MALDI-TOF mass spectrometric analyses (M–H negative ion mode) of (A) the multistep DNA-templated synthesis of 8a starting from 1a ($R$ is NHCH$_3$), (B) the translation of four templates (1a to 1d; $R$ is tryptamine) into four corresponding macrocycles (8a to 8d), and (C) step two of the translation of 65 templates (the 64 templates containing all possible combinations of codons complementing 2a to 2d, 3a to 3d, 4a to 4d, plus template 1e) into 65 corresponding macrocycles. (D) Analysis of the DNA-templated 65-member macrocyclic fumaramide library by denaturing PAGE (lanes a to c) and agarose gel electrophoresis (lanes d to g). Lane a, DNA-linked thiol reagent complementing the constant 5' region of all macrocycle template sequences; lane b, the 65-member library of template-linked macrocycles (8); lane c, the library after DNA-templated thiol addition, confirming the presence of the fumaramide group formed during macrocyclization; lane d, NlaIII digestion of PCR-amplified templates from the 65-member macrocycle library before selection; lanes e and f, NlaIII digestion of PCR-amplified templates from the 65-member macrocycle library after one and two rounds of selection for carbonic anhydrase affinity, respectively; lane g, NlaIII digestion of authentic PCR-amplified template 1e that directs the synthesis of 8e.
Hydrophobic Collapse in Multidomain Protein Folding

Ruhong Zhou,1,2* Xuhui Huang,2 Claudio J. Margulis,2 Bruce J. Berne1,2*

We performed molecular dynamics simulations of the collapse of a two-domain protein, the BphC enzyme, into a globular structure to examine how water molecules mediate hydrophobic collapse of proteins. In the interdomain region, liquid water persists with a density 10 to 15% lower than in the bulk, even at small domain separations. Water depletion and hydrophobic collapse occur on a nanosecond timescale, which is two orders of magnitude slower than that found in the collapse of idealized paraffin-like plates. When the electrostatic protein-water forces are turned off, a dewetting transition occurs in the interdomain region and the collapse speeds up by more than an order of magnitude. When attractive van der Waals forces are turned off as well, the dewetting in the interdomain region is more profound, and the collapse is even faster.

In the folding of globular proteins, it is often useful to picture the hydrophobic residues as being driven together by the action of water, in much the same way that droplets of oil would be driven together in water (1–4), but the presence of both a polar backbone and polar hydrophilic side chains complicates this picture. Most of our current understanding of hydrophobic collapse springs from studies on simple solutes (1–3, 5–18) or of model hydrophobic chains (19–22). The role of the hydrophobic interaction in the folding of peptide chains in water is unfortunately a complex problem. We have chosen to study a structurally simpler problem, the collapse of two-domain proteins, where the starting point is the already folded domains.

In a two-domain protein folding, we can probe the hydrophobic collapse and possible dewetting in the interdomain region when the two complementary domain surfaces (largely hydrophobic) approach each other. The relative stability of each individual domain and the comparable surface area of the interfacial region also make two-domain protein folding somewhat comparable to the previously studied collapse of idealized paraffin-like plates (3). Furthermore, by turning off various portions of the protein-water interaction, we can better understand the important features of collapse present in the case of proteins, but perhaps not in the collapse of paraffin-like plates or the aggregation of oil droplets.

We simulated, by molecular dynamics (MD), the hydrophobic interaction between the domains of a two-domain protein, the BphC enzyme (1dhy), which functions in degrading toxic polychlorinated biphenyls. One focus is on how the water molecules behave in the interdomain region when the multidomain protein folds (or collapses) into its final shape, after each individual domain has been formed, and how this behavior changes in response to changes in the protein-water interaction. The MD simulations were carried out with an all-atom model of both the

References and Notes

13. Synthesis and characterization details are available on Science Online.
19. Supported by NIH (National Institute of General Medical Sciences R01GM065865), the Office of Naval Research (N00014-03-1-0749), the Arnold and Mabel Beckman Foundation, the Searle Scholars Foundation (00-C-101), and the Alfred P. Sloan Foundation (BR-4141). Z.J.G. is a Bristol-Myers Squibb Graduate Research Fellow. B.N. and T.M.S. are NSF Graduate Research Fellows. J.B.D. is a National Defense Science and Engineering Graduate Research Fellow. We are grateful to the Bauer Center for Genomics Research for MALDI-TOF mass spectrometric analyses. G. Verdone for LC-MS instrument access, and DNA Software for assistance with codon screening. The rights to commercial development of DNA-templated synthesis have been licensed to Ensemble Discovery, a company for which D.R.L. is a consultant and shareholder.

*To whom correspondence should be addressed. E-mail: ruhong@us.ibm.com (R.Z.); berne@chem.columbia.edu (B.J.B.)